

Joint Graduate Presentation
Department of Microbiology

Nanogold technology in Microbiology

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- 2. Underlying principles
 - Physical properties of gold nanoparticles
- 3. Applications on biochemical methods
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1. Introduction

Current Applications of Nanotechnology:

Applications in Medical Sciences:

- 1. Antimicrobial (Silver)
- 2. Biomolecule detection (Gold)
- 3. Drug delivery (Carbon)

2. Underlying principles

- Physical properties of gold nanoparticles
- Detection is based on:
- For large nanogold particles ($>200\text{nm}$)
 - Precipitation of gold on target molecules
 - Net result: gold is seen on surface as deposits



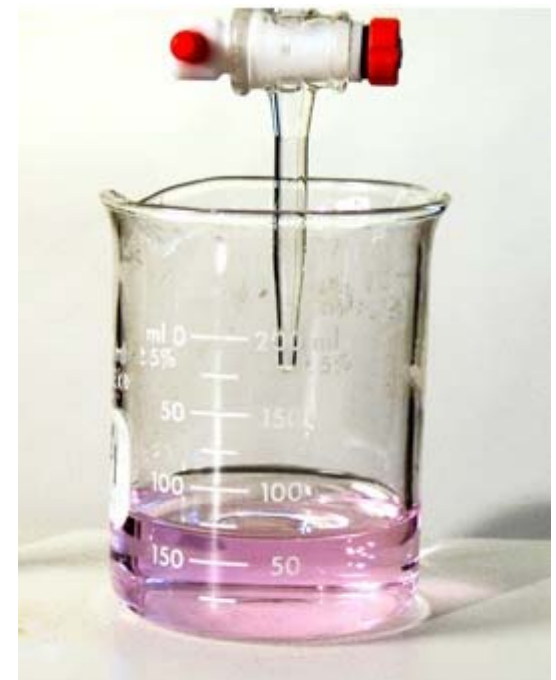
2. Underlying principles

- Large nanogold particles
 - Similar physical properties as its bulk material
 - Yellow in colour
 - The principle of detection is the precipitation of gold particles onto target
 - Have advantage of ease in detection (can be detected by a CCD camera alone)



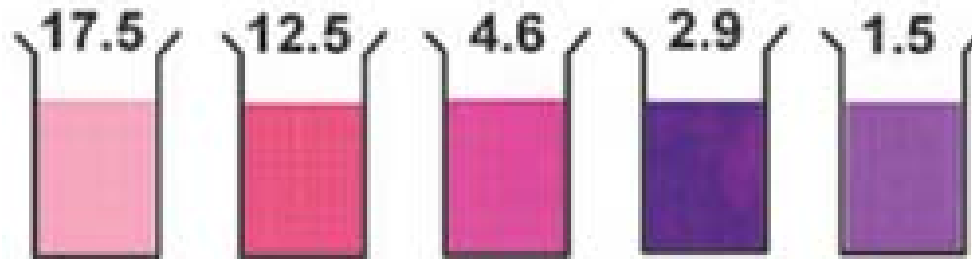
2. Underlying principles

- For small nanogold particles (2-100nm)
 - Change in absorption wavelength by plasmon resonance (i.e. colour is not gold anymore)
 - Net result: As a powerful red dye, or act as a distance dependent reporters



2. Underlying principles

- For small nanogold particles (2-100nm)
 - Give different physical properties than its bulk material
 - Change from red to blue when gold particles bring close together (i.e. distance dependent)



- Martin Holtzhauer, Michael Rudolph, Application of colloidal gold for characterization of supports used in size-exclusion chromatography, Journal of Chromatography A, Volume 605, Issue 2, 17 July 1992, Pages 193-198

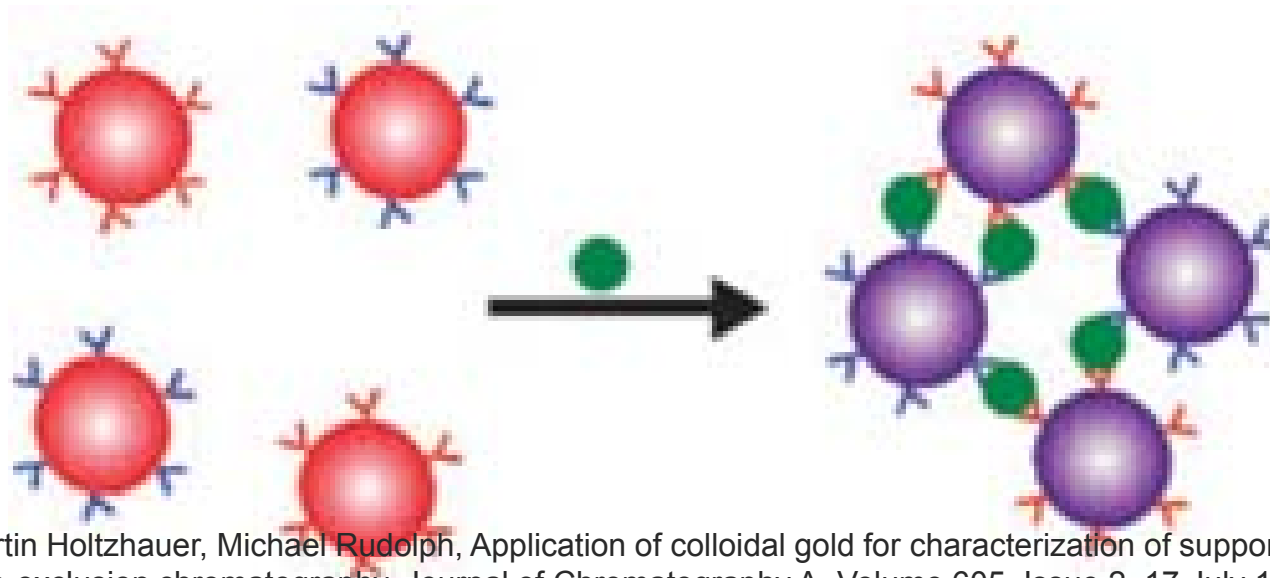
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3. Applications in biochemical methods

- Nanogold particles can be conjugated to antibodies, primers, or oligopeptides by interaction of thiol groups
- It can also conjugate to chemicals like aminodextran

3 [Applications in biochemical methods]

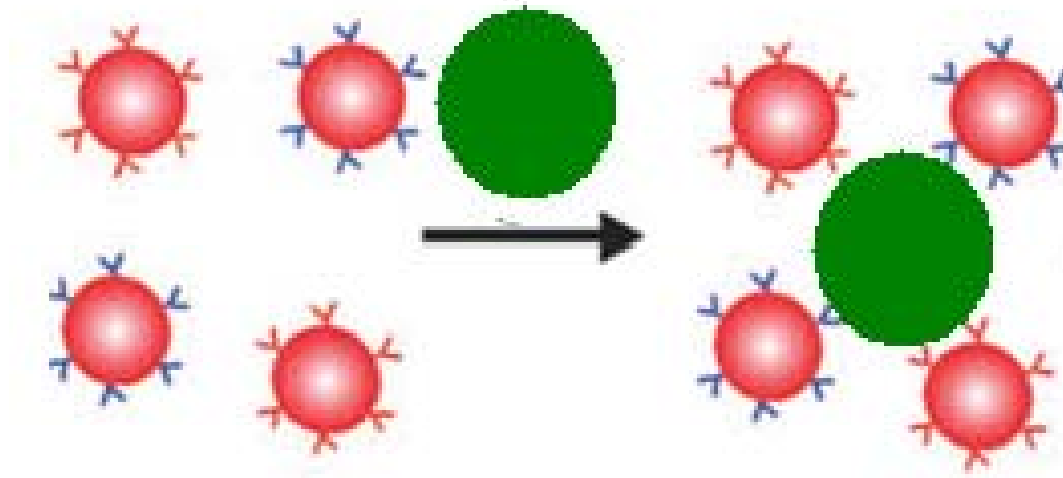
- Distance dependent detection of antigens
 - Conjugate to antibodies
 - **Antigen** + **Au-Antibodies** → Nanogold particles bring close together



- Martin Holtzhauer, Michael Rudolph, Application of colloidal gold for characterization of supports used in size-exclusion chromatography, Journal of Chromatography A, Volume 605, Issue 2, 17 July 1992, Pages 193-198

3 [Applications in biochemical methods]

- Large antigens...



- Solution: Use bigger size nanogold particles

3 Applications in biochemical methods

- Detection of enzymatic activity
 - E.g. Substrate specific protease:

- Cristian Guarise, Lucia Pasquato, Vincenzo De Filippis, and Paolo Scrimin
Gold nanoparticles-based protease assay
PNAS 2006 103: 3978-3982

AcNHCys(SAc)-peptide-Cys(SAc)OH
sequence specific for a protease

Incubate with protease then add to > 4 nm pink-red gold nanoparticles

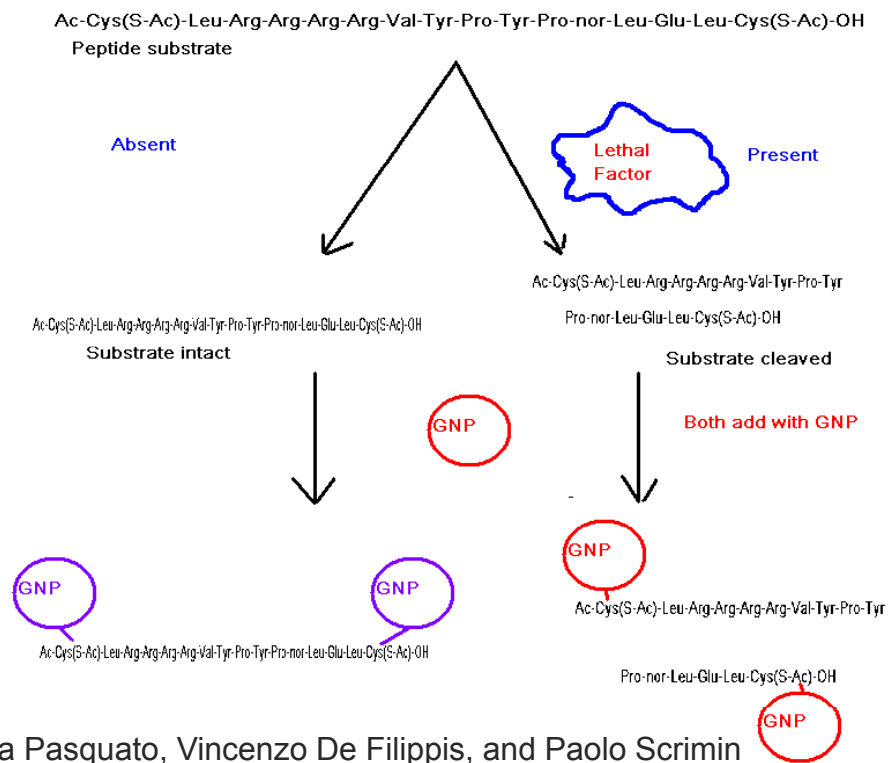
Color does not change:
protease is present
(cleaved peptide is unable to induce nanoparticle aggregation)

Color turns to blue-violet:
protease is absent
(uncleaved peptide induces nanoparticle aggregation)

Applications in Microbiology

■ Detection of enzyme activity

- Anthrax Lethal factor – Detection of *Bacillus anthracis*



- Cristian Guarise, Lucia Pasquato, Vincenzo De Filippis, and Paolo Scrimin
Gold nanoparticles-based protease assay
PNAS 2006 103: 3978-3982

3 [Applications in biochemical methods]

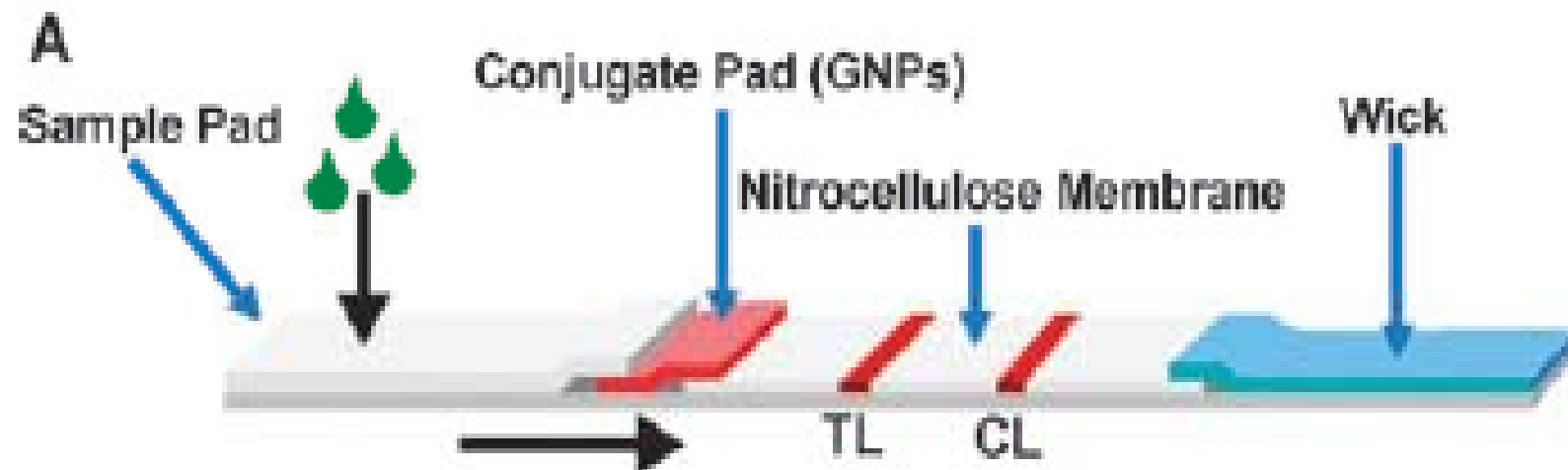
- Under investigations:
 - SNP detection by ligase reaction
 - Detection of amplified nucleic acid products

A colorimetric method for point mutation detection using high-fidelity DNA ligase
Jishan Li, Xia Chu, Yali Liu, Jian-Hui Jiang, Zhimin He, Zhiwei Zhang, Guoli Shen, and Ru-Qin Yu
Nucleic Acids Res. 2005; 33(19): e168

Eric Tan, Barbara Erwin, Shale Dames, Karl Voelkerding, and Angelika Niemz Isothermal DNA Amplification with Gold Nanosphere-Based Visual Colorimetric Readout for Herpes Simplex Virus Detection , Clin Chem 2007 53: 2017-2020

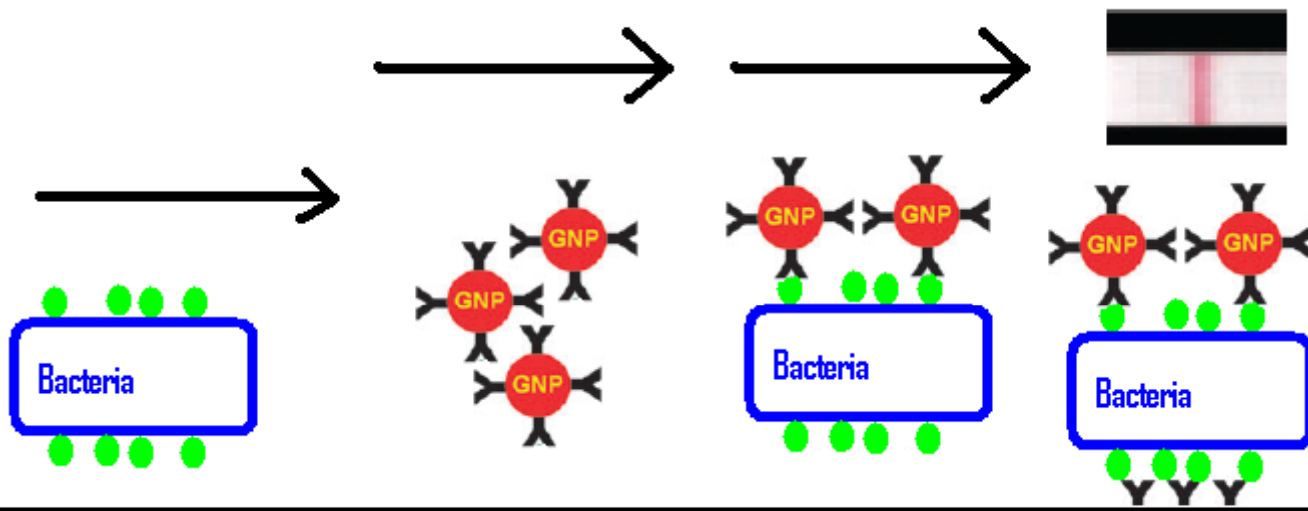
3 Applications in biochemical methods

- Nanogold as a dye
 - Lateral flow device



3 Applications in biochemical methods

- Nanogold as a dye
 - Lateral flow device

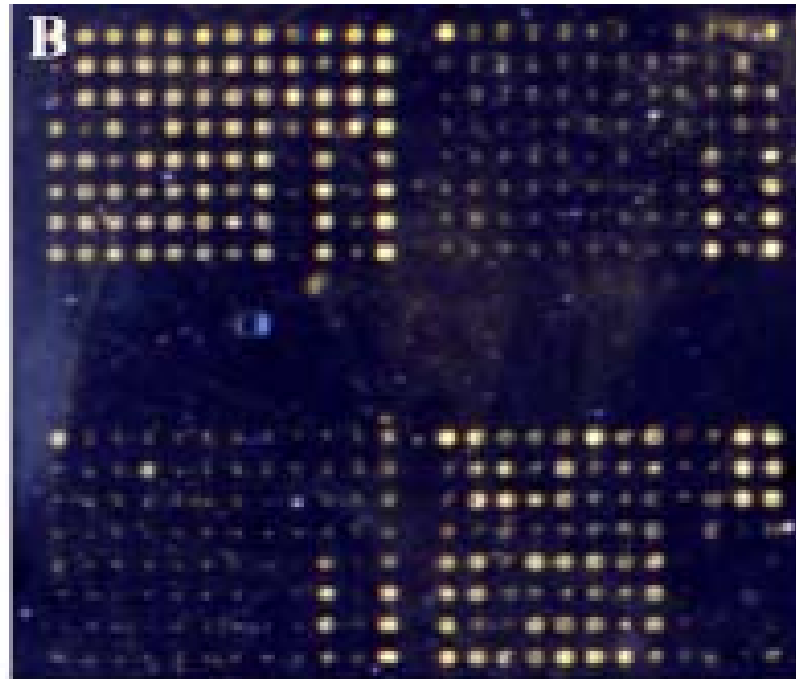
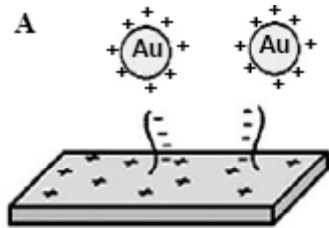


Applications in Microbiology

- Lateral flow devices:
 - Immunochromatographic test strip against *V. harveyi* in aquaculture
 - immunochromatographic assay for the detection of *S.aureus* in food
-
- Paisarn Sithigorngul, Sombat Rukpratanporn, Nilawan Pecharaburanin, Pornthip Suksawat, Siwaporn Longyant, Parin Chaivisuthangkura, Weerawan Sithigorngul, A simple and rapid immunochromatographic test strip for detection of pathogenic isolates of *Vibrio harveyi*, *Journal of Microbiological Methods*, Volume 71, Issue 3, December 2007, Pages 256-264
 - Su-Hua Huang, Gold nanoparticle-based immunochromatographic test for identification of *Staphylococcus aureus* from clinical specimens, *Clinica Chimica Acta*, Volume 373, Issues 1-2, November 2006, Pages 139-143

3 Applications in biochemical methods

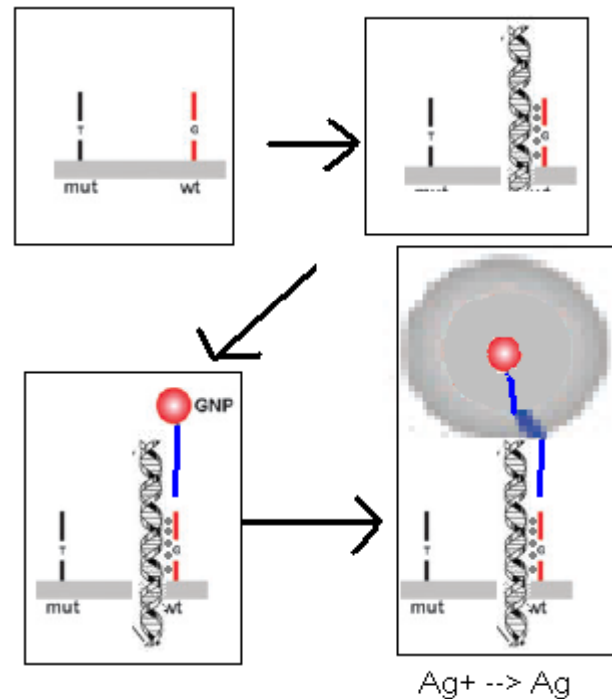
- Cationic 250nm nanogold particles
- Stronger attraction to Hybridized nucleotides (more negative)



- Y Sun, WH Fan, MP McCann, V Golovlev - Analytical biochemistry, 2005

3 Applications in biochemical methods

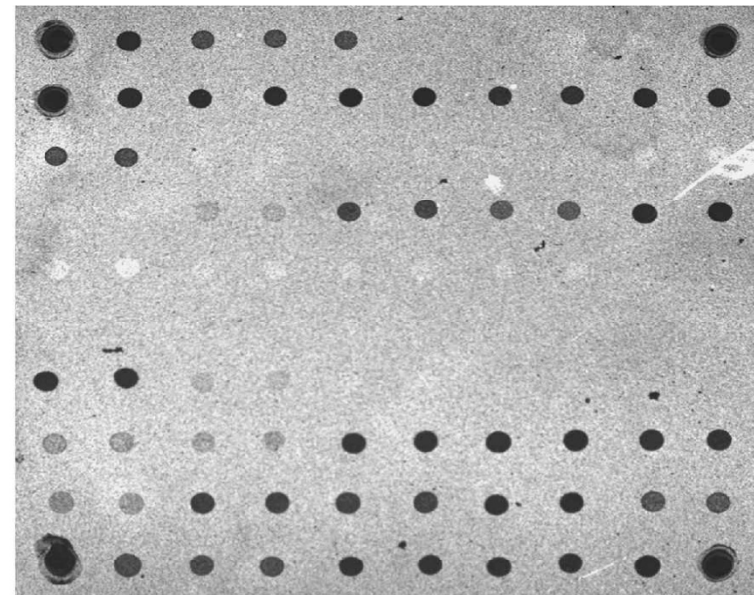
- Detection of unamplified sequence from genomic DNA, with silver enhancement



Applications in Microbiology

- Microarray for the screening of antibiotic resistance of *S.Aureus* and *M.tuberculosis*

91: POSITION MARKER	92: aphA3, 2	93: aphA3, 3	94: aphA3, 4	95: aphA3, 4	96: linA, 1	97: linA, 2	98: linA, 3	99: linA, 4	100: POSITION MARKER
81: POSITION MARKER	82: sat, 1	83: sat, 2	84: sat, 2	85: sat, 3	86: sat, 3	87: sat, 4	88: sat, 4	89: aphA3, 1	90: aphA3, 1
71: norA, 4	72: norA, 4	73: tst-1, 1	74: tst-1, 1	75: tst-1, 2	76: tst-1, 2	77: tst-1, 3	78: tst-1, 3	79: tst-1, 4	80: tst-1, 4
61: tetK, 3	62: tetK, 3	63: tetK, 4	64: tetK, 4	65: norA, 1	66: norA, 1	67: norA, 2	68: norA, 2	69: norA, 3	70: norA, 3
51: tetM, 2	52: tetM, 2	53: tetM, 3	54: tetM, 3	55: tetM, 4	56: tetM, 4	57: tetK, 1	58: tetK, 1	59: tetK, 2	60: tetK, 2
41: aphD, 1	42: aphD, 1	43: aphD, 2	44: aphD, 2	45: aphD, 3	46: aphD, 3	47: aphD, 4	48: aphD, 4	49: ---	50: ---
31: ermC, 4	32: ermC, 4	33: ermA, 1	34: ermA, 1	35: ermA, 2	36: ermA, 2	37: ermA, 3	38: ermA, 3	39: ermA, 4	40: ermA, 4
21: blaZ, 3	22: blaZ, 3	23: blaZ, 4	24: blaZ, 4	25: ermC, 1	26: ermC, 1	27: ermC, 2	28: ermC, 2	29: ermC, 3	30: ermC, 3
11: coA, 1	12: coA, 2	13: coA, 3	14: coA, 3	15: coA, 4	16: coA, 4	17: blaZ, 1	18: blaZ, 1	19: blaZ, 2	20: blaZ, 2
1: POSITION MARKER	2: mecA, 1	3: mecA, 2	4: mecA, 2	5: mecA, 3	6: mecA, 3	7: mecA, 4	8: mecA, 4	9: coA, 1	10: POSITION MARKER



- S. Monecke, I. Leube and R. Ehrlich, Genome Lett., 2003, 2, 106

Conclusion

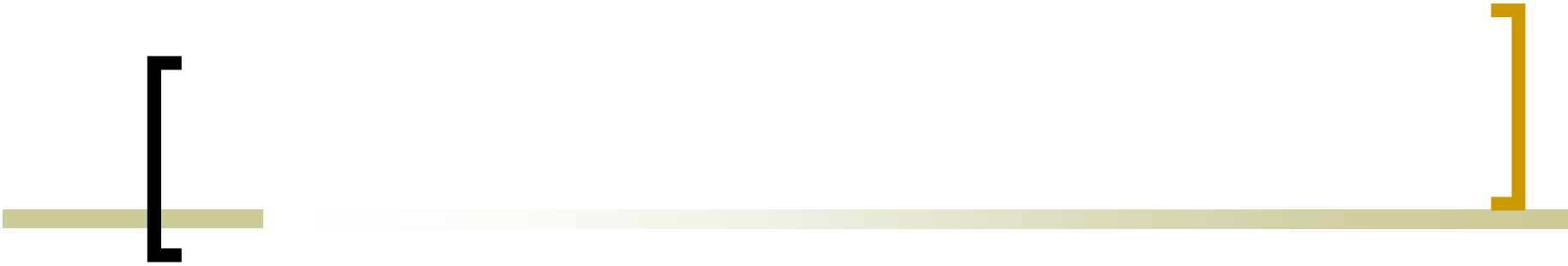
- When the size of the gold nanoparticles decrease, they possess different optical and physical characteristics
- Such characteristics can be utilized in many biochemical tests, and therefore useful in microbiology

[References]

- S. Monecke, I. Leube and R. Ehricht, *Genome Lett.*, 2003, 2, 106
- Paisarn Sithigorngul, Sombat Rukpratanporn, Nilawan Pecharaburanin, Pornthip Suksawat, Siwaporn Longyant, Parin Chaivisuthangkura, Weerawan Sithigorngul, A simple and rapid immunochromatographic test strip for detection of pathogenic isolates of *Vibrio harveyi*, *Journal of Microbiological Methods*, Volume 71, Issue 3, December 2007, Pages 256-264
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- Y Sun, WH Fan, MP McCann, V Golovlev - *Analytical biochemistry*, 2005

[References]

- A colorimetric method for point mutation detection using high-fidelity DNA ligase, Jishan Li, Xia Chu, Yali Liu, Jian-Hui Jiang, Zhimin He, Zhiwei Zhang, Guoli Shen, and Ru-Qin Yu, *Nucleic Acids Res.* 2005; 33(19): e168
- Eric Tan, Barbara Erwin, Shale Dames, Karl Voelkerding, and Angelika Niemz Isothermal DNA Amplification with Gold Nanosphere-Based Visual Colorimetric Readout for Herpes Simplex Virus Detection , *Clin Chem* 2007 53: 2017-2020
- Martin Holtzhauer, Michael Rudolph, Application of colloidal gold for characterization of supports used in size-exclusion chromatography, *Journal of Chromatography A*, Volume 605, Issue 2, 17 July 1992, Pages 193-198
- R Wilson - *Chemical Society Reviews*, 2008



End

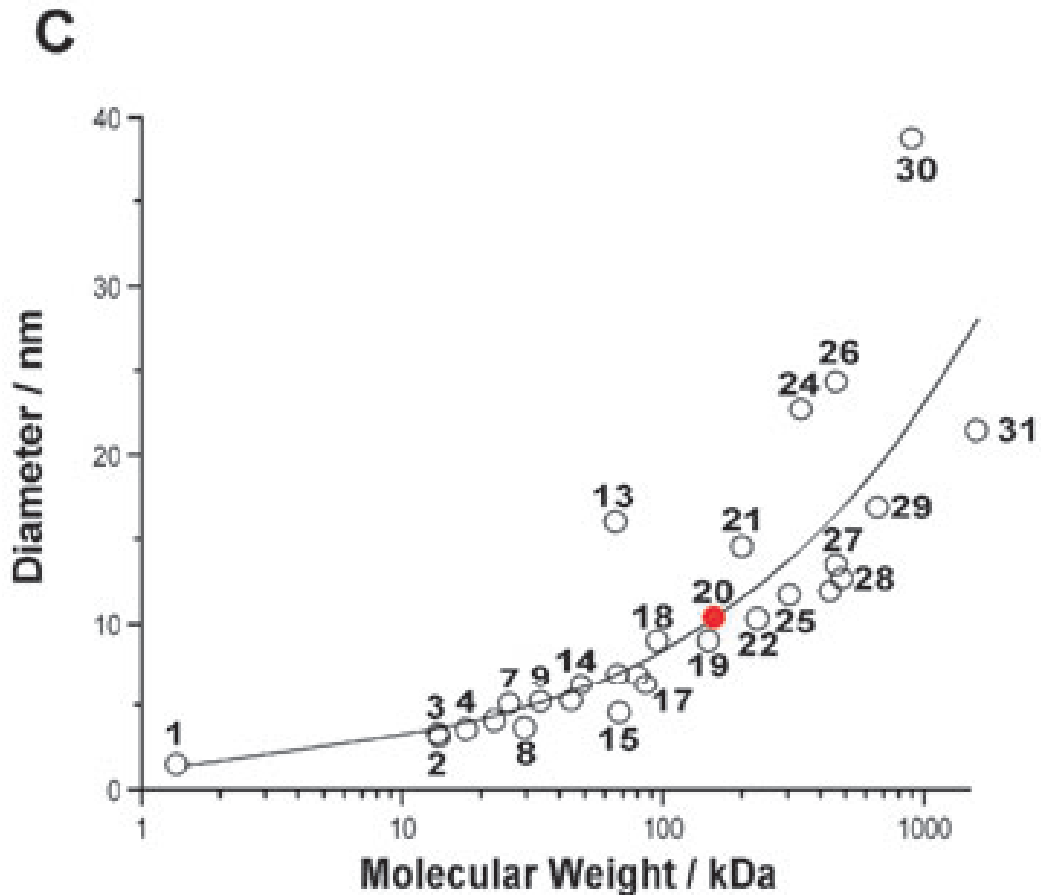
[Detection sensitivity]

- Protease assay: 25nM LF(by eye)
- Lateral flow device: 100 thousand cfu
- DNA probe: 10^{-11} mol target DNA (eye)

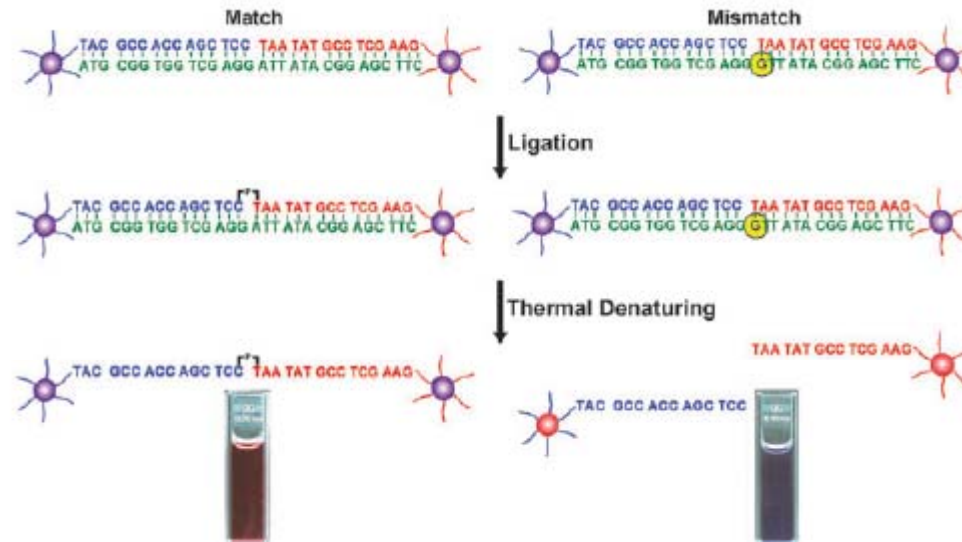
[Colour Intensity]

- Sybr safe: $7.3 \times 10^4 \text{M}$
- Nanogold (10nm): $\sim 1 \times 10^8 \text{M}$
- Nanogold (80nm): $5 \times 10^{10} \text{M}$

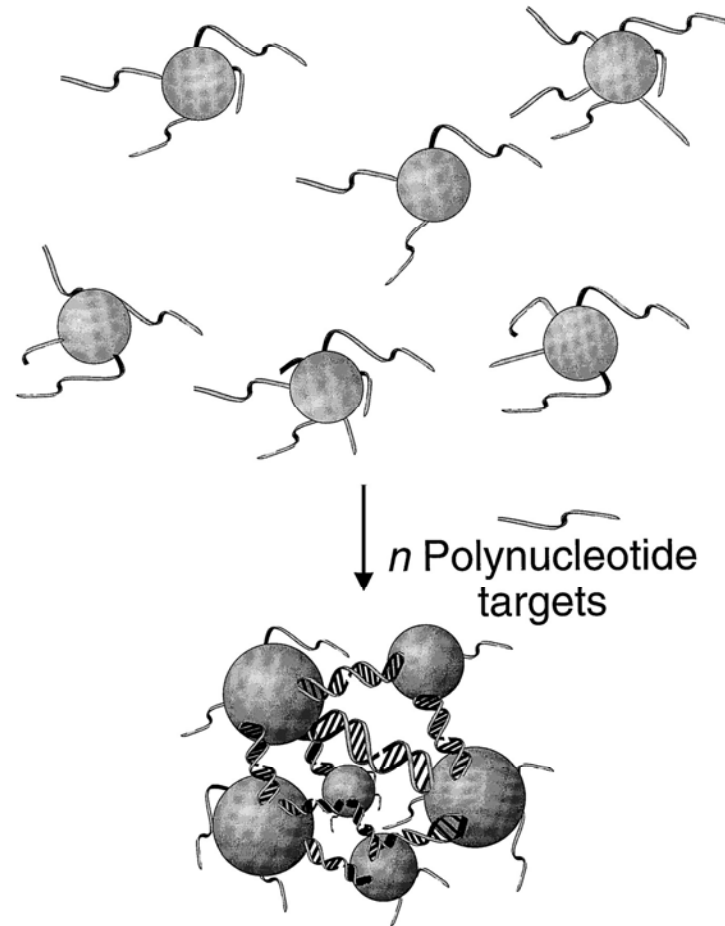
Size and max. distance



SNP detection



Amplified products



[DIY nanogold (10-20nm)]

- Use: conjugate to antibody
- 1. Disintergrate
- Aqua regia (nitro-hydrochloric acid) +
- Gold
- → Chloroauric acid

[DIY nanogold (10-20nm)]

- 2. Assemble
- Take 5.0×10^{-6} mol of HAuCl_4 , dissolve it in 19 ml of deionized water (the result should be a faintly yellowish solution).
- Heat it until it boils.
- Continue the heating and, while stirring vigorously, add 1 ml of 0.5% sodium citrate solution; keep stirring for the next 30 minutes.
- The colour of the solution will gradually change from faint yellowish to clear to grey to purple to deep purple, until settling on wine-red.
- Add water to the solution as necessary to bring the volume back up to 20 ml (to account for evaporation).

[Specificity and sensitivity]

